TECHNICAL NOTE

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JC virus genotyping offers a new means of tracing the origins of unidentified cadavers

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Abstract There has been no reliable means of tracing the origins of unidentified cadavers but the recent finding that JC virus (JCV) can serve as a means of elucidating human migrations suggested that this virus may also be useful to trace the origins of unidentified cadavers. DNA samples extracted from renal tissue and urine were used as the template for PCR amplification of a 610 bp region (IG region) of the viral genome. We detected JCV DNA in 45% of the renal samples and in 33% of the urine samples and was detectable even 10 days after death. The sequences of the amplified IG regions could be used to determine the genotypes. We conclude that the JC virus genotype is a new marker useful for tracing the origins of unidentified cadavers.

Keywords Forensic medicine · Unidentified cadaver · JC virus · Phylogenetic analysis · Genotypes · Geographic origin

Introduction

In spite of the progress in the methods of scientific investigation, the increase in the number of unidentified cadavers is a growing problem throughout the world [1, 2]. Recently, the relationships between several human DNA markers and human populations have been investigated [3, 4, 5], but these markers were of little use to elucidate the geographic origins of unidentified cadavers. The recent finding that JC virus (JCV) can serve as a means of tracing human migrations [6, 7] suggested that this virus may also be useful to elucidate the origins of unidentified

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cadavers. JCV asymptomatically infects most humans during childhood [8, 9] and the same viral strain persists in the kidneys throughout life [10, 11, 12]. JCV DNA can be classified into at least 12 genotypes that occupy distinct domains in different parts of the world [6, 13] (Table 1). Here, we report that a 610 bp region (IG region) of the JCV genome [14], usually used for the identification of JCV genotypes, can efficiently be amplified by PCR from the kidneys or urine of cadavers, and that the sequences of the amplified IG regions can be used to determine the genotypes.

Materials and methods

Subjects

A total of 64 renal tissue and 36 urine samples were obtained during forensic autopsies in our department. The origins of the cadavers were all known.

JCV genotype ^a	Geographic region where an indi- cated genotype is mainly detected ^b
EU (Types 1 and 4)	Europe, Mediterranean areas
B1-c (Type 2B)	Europe
Af1 (Type 6)	Central and western Africa
Af2 (Type 3)	Africa, western Asia
Af3	Central Africa
B1-a	China
B1-b (Type 2D)	Central and western Asia
B1-d	Saudi Arabia, Greece
B2	India, Mauritius
CY	Northeastern Asia
SC (Type 7)	Southeastern Asia, southern China
MY (Types 2A and 2C)	Japan, South Korea

^aGenotypes designated by Sugimoto et al. [6] and Guo et al. [13] are shown, and those designated by Jobes et al. [32] are shown in parentheses. ^bDomains are indicated according to Sugimoto et al. [6] and Guo et

^bDomains are indicated according to Sugimoto et al. [6] and Guo et al. [13]. Major JCV genotypes in the New World: EU, B1-c, Af1, Af2, MY [7, 32].

Extraction of DNA from kidney and urine

A 10×10×2 mm slice was excised from three different medullar regions in each of the bilateral kidneys. DNA was extracted as described previously [15]. Extraction of DNA from urine was also performed essentially as described [15].

Amplification of the IG region

Amplification of the 610 bp IG region was conducted using Pwo DNA polymerase with proof-reading activity (Boeringer Mannheim, Germany) as described [16].

Sequencing

The PCR-amplified fragments were cloned into pBluescript II SK(+) (Stratagene, La Jolla, Calif.) as described [12]. Recombinant plasmids were sequenced using an autosequencer ABI 3700 (Perkin-Elmer, Foster City, Calif.).

Phylogenetic analysis

Neighbor-joining phylogenetic trees [17] were constructed using the CLUSTAL W program [18].

Results

JCV DNA was usually detected only in part of the tissue specimens derived from the same cadaver (Table 2). JCV DNA was detected in both kidneys in 15 cases, in the left kidney only in 7 cases, and in the right kidney only in 8 cases. Thus, JCV DNA detection had no bias towards one of the bilateral kidneys. JCV DNA was efficiently detected in older age groups (Table 3) and was also efficiently detected up to 72 h after death (Table 4). We also attempted to detect JCV DNA in cadaveric urine specimens but this was less efficient than from kidneys (Table 4). We examined the correlation between the JCV genotype and the origins of cadavers. JCV genotypes detected in the kidney or urine specimens from cadavers are shown for each geographic region in Table 5. Two major JCV genotypes, CY and MY were detected in Japanese cadavers, CY was detected in all cadavers originating from northern China and CY and MY were detected in cadavers from Korea.

Discussion

In this study, we obtained the following findings as to the detection of the JCV DNA from the kidney of cadavers:

- 1. JCV DNA was detected in both kidneys
- 2. JCV DNA was efficiently detected from elderly cadavers aged 25 years or more at death
- 3. JCV DNA was not detected at all sites in the kidneys.

The last two findings were similar to those in living subjects [11, 15, 19, 20, 21], but the first finding has not been reported in studies using living subjects.

From the findings in this study we concluded that the JC virus genotype is a new marker useful for tracing the origins of unidentified cadavers. Many genetic markers, including mtDNA, Y chromosome, and microsatellites, have been used to analyse the relationships among various human populations. However, although these markers are useful in population-based studies, they hardly provide reliable information on the ethnic origin of individuals.

Table 2 Detection of JCVDNA in various areas of bilat-		Not detected	Detected	No. of positive samples/no. of samples analysed (%)					
eral kidneys				1/6	2/6	3/6	4/6	5/6	6/6
	No. of cadavers	35	29 (100)	7 (24)	6 (21)	9 (31)	5 (17)	1 (3)	1 (3)

Table 3 Detection of JCV DNA from cadaveric kidneys and urine in various age groups	Tissue	1	No. of positive cadavers/no. of cadavers examined (%) in age group (years)			
		0–25	26–50	51-75	>76	
$^*P < 0.05$ (vs. urine in the same age group).	Kidney Urine	0/5 (0) 1/5 (20)	11/32 (34) 4/13 (31)	14/21 (67)* 5/16 (31)	4/6 (67) 1/2 (50)	29/64 (45)* 11/36 (31)

Table 4	Detection of JCV	DNA in	the cadaveric	kidneys and	urine at	various	post-mortem times

No. of positive ca	No. of positive cadavers/no. of cadavers examined (%) at time post-mortem (hours)				
0–24 h	25–48 h	49–72 h	>73 h		
19/44 (43)	6/11 (55)	3/6 (50)	$\frac{1/3}{1/1}$ (33) ^a	29/64 (45) 11/36 (31)	
	0–24 h	0–24 h 25–48 h 19/44 (43) 6/11 (55)	0-24 h 25-48 h 49-72 h 19/44 (43) 6/11 (55) 3/6 (50)	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	

^aJCV DNA was detected in a single cadaver 7 days after death.

^bJCV DNA was detected in a single cadaver 10 days after death.

 Table 5 JCV genotypes in cadavers of various origins

Origins of cadavers	No. of JCV DNA positive cadavers	No. of cadavers with genotype		
		CY	MY	
Japan				
Southwestern area ^a	4	3	1	
Northeastern area ^a	5	0	5	
Intermediate area ^a	26	15	11	
Out side Japan				
Northern China ^b	3	3	0	
South Korea	2	1	1	

^aAccording to Kitamura et al. [31] Japan was divided into 3 areas, southeastern, northeastern and intermediate areas.

^bAccording to Guo et al. [13] China was divided into 2 areas, northern and southern areas.

Therefore, at present the JCV genotype is the most useful marker that can be used to elucidate the origins of unidentified cadavers. The JCV marker cannot precisely locate the origin of cadavers, but can restrict the area to be investigated to trace the origin. The information concerning the JCV genotypes existing in cadavers could be one of the most important individual elements to be considered in the investigation of unidentified cadavers. Furthermore, if the geographic distribution of JCV genotypes is clarified in detail in the future, the application of the current method will be expanded. In combination with other methods [22, 23, 24, 25, 26, 27, 28, 29, 30], this new method should make the investigation of unidentified cadavers both more efficient and economical.

To emphasise the usefulness of our method, we present the following examples in which the detection of JCV genotypes was helpful to identify the cadavers. In the first case, a cadaver was found in Tokyo, which was apparently Japanese, but there were no belongings which could have been useful to determine the geographic origin of the cadaver. We detected the MY genotype in this cadaver and based on this information, an investigation to search for the origin of this cadaver was first performed in the northeast area of Japan, where MY is more predominant [31]. This cadaver was identified among the missing in the northeast area of Japan. In the second case the SC genotype which is rare in Japan was detected in a cadaver found in Tokyo. Since SC is rarely found in Japanese [6, 31], the investigation was extended abroad to southern China and southeastern Asia where this genotype is predominant. This cadaver was identified among the missing persons from southern China.

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References

- 1. Ministry of Health and Welfare Japan (2000) Vital statistics Japan, vol. 3
- 2. Cattaneo C, Ritz-Timme S, Shutz HW, et al, (2000) Unidentified cadavers and human remains in the EU: an unknown issue. Int J Legal Med 113:N2–N3
- Rohl A, Brinkmann B, Forster L, Forster P (2001) An annotated mtDNA database. Int J Legal Med 115:29–39
- Brace CL (1995) Region does not mean race reality versus convention in forensic anthropology. J Forensic Sci 40:171–175
- 5. Cann RL, Stoneking M, Wilson AC (1987) Mitochondrial DNA and human evolution. Nature 325:31–36
- 6. Sigimoto C, Kitamura T, Guo J, et al, (1997) Typing of urinary JC virus DNA offers a novel means of tracing human migrations. Proc Natl Acad Sci USA 94:9191–9196
- Agostini HT, Yanagihara R, Davis V, Ryschkewitsch CF, Stoner GL (1997) Asian genotypes of JC virus in Native Americans and in a Pacific Island population: markers of viral evolution and human migration. Proc Natl Acad Sci USA 94:14542– 14546
- Padgett BL, Walker DL (1973) Prevalence of antibodies in human sera against JC virus, an isolate from a case of progressive multifocal leukoencephalopathy. J Infect Dis 127:467–470
- 9. Padgett BL, Walker DL (1976) New human papovaviruses. Prog Med Virol 22:1–35
- 10. Chesters PM, Heritage J, McCance DJ (1983) Persistence of DNA sequences of BK virus and JC virus in normal human tissues and in diseased tissues. J Infect Dis 147:676–684
- Tominaga T, Yogo Y, Kitamura T, Aso Y (1992) Persistence of archetypal JC virus DNA in normal renal tissue derived from tumor-bearing patients. Virology 186:736–741
- 12. Kitamura T, Sugimoto C, Kato A, et al, (1997) Persistent JC virus (JCV) infection is demonstrated by continuous shedding of the same JCV strains. J Clin Microbiol 35:1255–1257
- 13. Guo J, Suchimoto C, Kitamura T, et al, (1998) Four geographically distinct genotypes of JC virus are prevalent in China and Mongolia: implications for the racial composition of modern China. J Gen Virol 79:2499–2505
- 14. Ault GS, Stoner GL (1992) Two major types of JC virus defined in progressive multifocal leukoencephalopathy brain by early and late coding region DNA sequences. J Gen Virol 73: 2669–2678
- 15. Kitamura T, Aso Y, Kuniyoshi N, Hara K, Yogo Y (1990) High incidence of urinary JC virus excretion in nonimmunosuppressed older patients. J Infect Dis 161:1128–1133
- 16. Kunitake T, Kitamura T, Guo J, et al, (1995) Parent-to-child transmission is relatively common in the spread of the human polyomavirus JC virus. J Clin Microbiol 33:1448–1451
- 17. Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol Biol Evol 4:406–425
- 18. Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position specific gap penalties and weight matrix choice. Nucleic Acids Res 22: 4673–4680
- Kitamura T, Kunitake T, Guo J,Tominaga T, Kawabe K, Yogo Y (1994) Transmission of the human polyomavirus JC virus occurs both within the family and outside the family. J Clin Microbiol 32:2359–2363
- Agostini HT, Ryschkewitsch CF, Stoner GL (1996) Genotype profile of human polyomavirus JC excreted in urine of immunocompetent individuals. J Clin Microbiol 34:159–164
- 21. Chang D, Sugimoto C, Wang M, Tsai RT, Yogo Y (1999) JC virus genotypes in a Taiwan aboriginal tribe (Bunun): implications for its population history. Arch Virol 144:1081–1090
- 22. Schmeling A, Reisinger W, Loreck D, Vendura K, Markus W, Geserick G (2000) Effects of ethnicity on skeletal maturation: consequences for forensic age estimations. Int J Legal Med 113:253–258

- 23. Ritz-Timme S, Cattaneo C, Collins MJ, Waite ER, Shutz HW, Kaatsch HI, Borrman HIM (2000) Age estimation: the state of the art in relation to the specific demands of forensic practice. Int J Legal Med 113:129–136
- 24. Wahl J, Graw M (2001) Metric sex differentiation of the pars petrosa ossis temporalis. Int J Legal Med 114:215–223
- 25. Pfeiffer H, Brinkmann B, Huhne J, Rolf B, Morris AA, Steighner R, Holland MM, Foster P (1999) Expanding the forensic German mitochondrial DNA control region database: genetic diversity as a function of sample size and microgeography. Int J Legal Med 112:291–298
- 26. Pfeiffer H, Forster P, Ortmann C, Brinkmann B (2001) The results of an mtDNA study of 1200 inhabitants of a German village in comparison to other Caucasian databases and its relevance for forensic casework. Int J Legal Med 114:169–172
- 27. Lutz S, Weisser H-J, Heizmann J, Pollak S (1999) Location and frequency of polymorphic positions in the mtDNA control region of individuals from Germany (Erratum). Int J Legal Med 112:205–209

- 28. Parson W, Parsons TJ, Scheithauer R, Holland MM (1998) Population data for 101 Austrian Caucasian mitochondrial DNA d-loop sequences: application of mtDNA sequence analysis to a forensic case. Int J Legal Med 111:124–132
- 29. Grignani P, Peloso G, Fattorini P, Previdere C (2000) Highly informative Y-chromosomal haplotypes by the addition of three new STRs DYS437, DYS438 and DYS439. Int J Legal Med 114:155–129
- 30. Iida R, Tsubota E, Matsuki T (2001) Identification and characterization of two novel human polymorphic STRs on the Y chromosome. Int J Legal Med 115:54–56
- 31. Kitamura T, Sugimoto C, Ebihara H, et al, (1998) Peopling of Japan as revealed by genotyping of urinary JC virus DNA. Anthropol Sci 106:311–325
- 32. Jobes DV, Chima SC, Ryschkewitsch CF, Stoner GL (1998) Phylogenetic analysis of 22 complete genomes of the human polyomavirus JC virus. J Gen Virol 79:2491–2498